Review Article - The Use of Zinc Oxide (Zno)Nano Technology as An Alternative To Antibiotics in Veterinary Medicine Fields

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Abstract

Zinc oxide (ZnO) is used in place of an alternative or toward reduce antibiotics, and it has stayedgenerally used in wound healing and numerous skin conditions. This is why nanotechnology has opened up significant, advanced, or new expansions in applications in nearly all disciplines of the general sciences, including veterinary and animal sciences. ZnO nanoparticles (NPs) consume recently gained wide devotion due to their single and distinct properties. ZnO NPs may have many applications due to their anti-bacterial, neoplastic, and wound-healing properties. Lots of recent research into the field of ZnO NPs and their potential for application in the fields of veterinary medicine.

Key words: Nanotechnology a. Zinc nanoparticles, antibacterial motion

Introduction

Nanotechnology is at the front of studyin addition has enormous possible to transform the livestock subdivision. Through the arrival of this evolving field, a widespread variety Nanoparticles with interesting properties are manufactured and used in a wide range of applications. [1] The amount of manufactured nanoparticles will increase to 5,000,000 tons by 2020. Metal oxide nanoparticles are commonly used in steroids, sensors, environmental treatments, personal care products, and zinc oxide (ZnO).has been widely used among metal nanoxides due to its antimicrobial, anti-biotic and neoplastic activities [2,3,4,5].

Commonly used ZnO is as a pseudo-substance in cosmetic lotion because it is used in h6 for h6 for h6 for rn to absorb UV rays and to block their properties. It is also used as an astringent medicine for bleeding ,ZnO NPs have recently gained attention due to their various properties. ZnO NPs may have many potential tools in veterinary science because of their antibacterial, neoplastic, wound-healing and vascular possessions. [7].

In animals, the inability of drugs to target meningitis, tumor types, and diseases caused by many pathogens inside the womb such as viruses and microorganisms (Brucella, Rickettsia, Chlamydia, etc.) and fungi (Histoplasma capsulatum and Cryptococcus neoformans) have already been treated. it's hard. Place. To treat this crisis, the beneficialagent'sobligation penetrate confidential the cells and be able to irritated the blood-brain blockade. This remains not likelybycurrent large-molecular therapies, then nanoparticles can showin contradiction of intracellular pathogens then brain tumors because of their minormass [8].

Antimicrobial properties

Antibiotic use in large quantities has limited therapeutic effect because their continued use leads to antimicrobial resistance [9]. New alternatives in modern science, particularly inanimate nanoparticles, have newlyadded attention. In metal oxide nanoparticles, Zeno NPs are commonly used for their antimicrobial properties. At the nanoscale, Metal oxide acquires certain properties that depend on mass, biochemical composition and surface chemistry. ZnO has been found to decrease the activity of a wide variety of bacteria, and the use of NPs significantly enhances the antibacterial property. The precise mechanism of antibacterial action is through binding of ZnO NPs to protein molecules to stop the cellular metabolism of bacteria and ultimately lead to the death of the microbes [10, 11, 12–14]

The size of nanoparticles has a great influence comparative studies on the antibacterial effect of together microns and nanoparticles of ZnO have shown that the antibacterial effect of ZnO NPs is high, due to the increase in cell membrane and cytoplasm, ZnO NPs may be perfect to replace particular of the existing antibiotics, ZnO NPs have additionalmarked antibacterial properties because of their minor their mass and height ratio of shallow to volume. (15-19).

ZnO NPs may be applied to animals in conditions such as mastitis, where they may be inaccessible by conventional antibiotics. Mastitis is usually Staphylococcus, Streptococcus and E. coli a disease that affects high-yielding animals. The economic result of mastitis include milk production, drug abuse and veterinary services [20,21]. Mastitis it is problematic to treat because of antibiotic fighting. The incidence of biofilms within intestinal tissues renders bacteria impermeable to physiological, chemical, and even innate immune mechanisms S. Aurius and H. K ZNO NP. It has been found to be effective against Escherichia coli, the management of staphylococcal mastitis in animals is already challenging because of the tendency of intracellular organisms. Since the nanostructures are capable to penetrate into cells, ZnO NPs can be used in staphylococcal mastitis. [24, 25].

In record cases, the pH of mastitis milk is alkaline in natural. Anti-bacterial at high concentrations in an alkaline medium would be the right choice to deal with this condition. Whereas, it was found that the antibacterial activity of Zn NPs against corona was higher at basic pH [26]. Escherichia coli is a Gram-negative bacterium accountablein place of coliform mastitis in ruminant and coliform granulomas in birds. ZnO NP in the Mail. It has strong antibacterial activity in contradiction of Escherichia coli [27-30]. Fall typhoid is a columnar infection caused by Salmonella galinarum in chickens worldwide, and ZnO NPs have been found to be effective in contradiction of Salmonella typhi and Staphylococcus aureus [31-34].

Listeria disease is a common disease in veterinary medicine, also known as vertigo, and results from the single root cause of Listeria disease, which is characterized by encephalitis, septicemia, and miscarriage in abortion. ZnO NPs at various concentrations L were effective against monocytogens [35]. Pseudomonas aeruginosa is a rod-shaped Gram-negative organism that selects to live in humid areas of the body such as the respiratory and gastrointestinal tracts in animals and humans [36,37]. It is intrinsically resistant to a wide range of antimicrobials, such as drug-flow mechanisms and the presence of purines due to L-lactam, tetracycline, and chloramphenicol. Biofilm formed by this organism is another antimicrobial inhibitor, ZnO NPs was found. Most effective against Pseudomonas [38,39].

Anthrax bacilli, the spore-producing bacteria, infects anthrax in warm-blooded animals. The production of anaerobic spore microbes causes many diseases in field animals such as tetanus, food poisoning, enterotoxicity, black quarters, and brakes. The spores shaped by these organisms live in the atmosphere for a long time and ripen into vegetative bacilli in a favorable state.

These germs are very impermeable to top temperatures and pressure. While ZnO NPs were establish to be operative contradiction of bacterial spores, ZnO NPs were synthesized from a plant basis that demonstrated antimicrobial activity against different pathogens compared to the chemical Znn NPs. Activity increased with increasing dose and treatment time [40, 41].

Anti-cancerNanotechnology

Among a current cancer actionplans, chemotherapy agents are usually used in medical patients. Presently, the current anti-neoplastic drugs are not very actual in treating patients because of absence of selective poisonousness. The indiscriminate use of antineoplants can cause adverse effects such as bone marrow suppression, neurotoxicity, and cardiomyopathy [42, 43]. Therefore, great emphasis has been placed on developing new anti-cancer agents can differentiate cancer cells from normal cells. It opened up a new nanotechnology horizons aimed at treating and diagnosing cancerm Nanoparticles be able to be used as a vehicle designed forunder attack delivery to tumor sites , the main advantage is that it can be absorbed by exact cells and the center can be absorbed along with its external chemistry. [44-47].

ZnO NP can be applied for clinical purposes and sometimes in curative cases in oncological conditions, and is usually used to treat lymphomas, melanomas, gastrointestinal tumors and sarcoidosis. ZnO NPs act on suitable concentrations of cialocaine with different concentrations as TNF- α , IFN-IF, and IL-12 in vitro and in vivo (lung). Cytokines encouraged by nanoparticles canistersimplify effective protective activities by removing appropriate cytokine profiles to enhance immunity by Th1 [48]. ZnO NPs have been shown to possess in height selective cytotoxicity, with a preference for killing tumor cells with minimal toxicity in usual primary immune cells, and diagnostic tools based on ZnO NPs aimed atdiscovery of little levels of biomarkers intended for cancer diagnosis are useful in [49,50].

Tumors of the right system are the most common oncological disorders in pets , harmony was established between melanoma and ultraviolet radiation, In dogs, highly penetrating ionizing radiation causes leukemia and lung cancer, In hematopoietic tumors, lymphoma is more common in dogs and manifests itself as general lymphadenopathy [51]. Canine genital tumor, similarly known as an adhesive tumor, spreads from burned skin or mucous membrane during sexual intercourse and licking [52]. In addition to the facial area, it can also occur on the mucous membranes of the skin and anus, metastasis with this tumor is rare, but local lymph nodes and vital organs have been stated [53,54].

Mammary gland tumors are most often seen in dogs [55]. Equally, it is the most common melanoma of horse, mule and donkey, the infection is closely related to bovine papillomavirus types 1 and 2. In these settings, the use of ZnO NP can be detected in both clinical and therapeutic methods.[56-59].

References

- 1. Zhao J, Castranova V (2011) Toxicology of nanomaterials used in nanomedicine. J Toxicol Environ Health B Crit Rev 14: 593-632.
- 2. *Kumar C* (2006) *Nanomaterials: Toxicity, Health and Environmental Issues: Nanotechnologies for the Life Sciences (1stedn.) Wiley-VHC, Weinheim, Germany.*
- 3. Sun T, Yan Y, Zhao Y, Guo F, Jiang C (2012) Copper oxide nanoparticles induce autophagic cell death in A549 cells. PLoS One 7: e43442.
- 4. **Petchthanasombat C, Tiensing T, Sunintaboon P (2012)** Synthesis of zinc oxideencapsulated poly(methyl methacrylate)-chitosan core-shell hybrid particles and their electrochemical property. J Colloid Interface Sci 369: 52-57.
- 5. Becheri A, Durr M, Nostro PL, Baglioni P (2008) Synthesis and characterization of zinc oxide nanoparticles: application to textiles as UV-absorbers. J Nanopart Res 10: 679-689.
- 6. Sweetman SC (2005) The Complete Drug Reference (34thedn.) Pharmaceutical Press, London, UK.
- 7. *Manuja A, Kumar B, Singh RK (2012)* Nanotechnology developments: opportunities for animal health and production. Nanotechnology Development 2: 17-25.
- 8. **Baquero F, Coque TM, de la Cruz F** (2011) Ecology and evolution as targets: the need for novel eco-evo drugs and strategies to fight antibiotic resistance. Antimicrob Agents Chemother 55: 3649-3660.
- 9. Söderberg TA, Sunzel B, Holm S, Elmros T, Hallmans G, et al. (1990) Antibacterial effect of zinc oxide in vitro. Scand J PlastReconstr Surg Hand Surg 24: 193-197.
- 10. Jin T, Sun D, Su JY, Zhang H, Sue HJ (2009) Antimicrobial efficacy of zinc oxide quantum dots against Listeria monocytogenes, Salmonella Enteritidis, and Escherichia coli O157:H7. J Food Sci 74: M46-52.
- 11. **Yamamoto O (2001)** Influence of particle size on the antibacterial activity of zinc oxide. Int J Inorg Mater 3: 643-646.
- 12. Dodd AC, McKinley AJ, Saunders M, Tsuzuki T (2006) Effect of particle size on the photocatalytic activity of nanoparticulate zinc oxide. J Nanopart Res 8: 43-51.
- 13. Zhang L, Jiang Y, Ding Y, Povey M, York D (2007) Investigation into the antibacterial behaviour of suspensions of ZnO nanoparticles (ZnO nanofluids). J Nanopart Res 9: 479-489.
- 14. Stoimenov PK, Klinger RL, Marchin GL, Klabunde KJ (2002) Metal oxide nanoparticles as bactericidal agents. Langmuir 18: 6679-6686.
- 15. Sawai J (2003) Quantitative evaluation of antibacterial activities of metallic oxide powders (ZnO, MgO and CaO) by conductimetric assay. J Microbiol Methods 54: 177-182.
- 16. Kim YS, Seo JH, Cha HJ (2003) Enhancement of heterologous protein expression in Escherichia coli by co-expression of nonspeci?cDNAbinding stress protein, Dps. Enzyme Microb Technol 33: 460-465.
- 17. Kirchner C, Liedl T, Kudera S, Pellegrino T, Muñoz Javier A, et al. (2005) Cytotoxicity of colloidal CdSe and CdSe/ZnS nanoparticles. Nano Lett 5: 331-338.
- 18. Moghaddam AB, Mohammadi A, Dinarvand R, Badraghi J, Atyabi F, et al. (2008)Bioelectrocatalysis of Methyldopa by Adsorbed Tyrosinase on the surface of modified glassy carbon with carbon nanotubes. Int J Electrochem Sci 3: 291.

- 19. He YT, Wan J, Tokunaga T (2008) Kinetic stability of hematite nanoparticles: the effect of particle sizes. J Nanopart Res 10: 321-332.
- 20. Clement JL, Jarrett PS (1994) Antibacterial silver. Met Based Drugs 1: 467-482.
- 21. Jones N, Ray B, Ranjit KT, Manna AC (2008) Antibacterial activity of ZnO nanoparticle suspensions on a broad spectrum of microorganisms. FEMS Microbiol Lett 279: 71-76.
- 22. Jiang J, Oberdörster G, Biswas P (2009) Characterization of size, surface charge and agglomeration state of nanoparticle dispersions for toxicological studies. J Nanopart Res 11: 77-89.
- 23. Fu G, Vary PS, Lin CT (2005) Anatase TiO2 nanocomposites for antimicrobial coatings. J Phys Chem B 109: 8889-8898.
- 24. Erskine RJ, Wagner S, DeGraves FJ (2003) Mastitis therapy and pharmacology. Vet Clin North Am Food AnimPract 19: 109-138.
- 25. Seegers H, Fourichon C, Beaudeau F (2003) Production effects related to mastitis and mastitis economics in dairy cattle herds. Vet Res 34: 475-491.
- 26. Olson ME, Ceri H, Morck DW, Buret AG, Read RR (2002) Biofilm bacteria: formation and comparative susceptibility to antibiotics. Can J Vet Res 66: 86-92.
- 27. Jalal R, Saliani M, Goharshadi EK (2011) Effects of pH and concentration on antibacterial activity of ZnO nanofluids against Staphylococcus aureus. National Iranian New Chemistry Congress, Shiraz, Iran, pp. 885-890.
- 28. Bajpai KS, Chand N, Chaurasia V (2012) Nano zinc oxide-loaded calcium alginate films with potential antibacterial properties. Food Bioprocess Techn 5: 1871-1881.
- 29. Tayel AA, El-tras WF, Moussa S, El-baz AF, Mahrous H, et al. (2011) Antibacterial action of zinc oxide nanoparticles against foodborne pathogens. J Food Safety 31: 211-218.
- 30. Brayner R, Ferrari-Iliou R, Brivois N, Djediat S, Benedetti MF, et al. (2006) Toxicological impact studies based on Escherichia coli bacteria in ultrafine ZnO nanoparticles colloidal medium. Nano Lett 6: 866-870.
- 31. Padmavathy N, Vijayaraghavan R (2008) Enhanced bioactivity of ZnO nanoparticles an antimicrobial study. Sci Technol Adv Mater 9: 1-7.
- 32. Wang C, Liu LL, Zhang AT, Xie P, Lu JJ, et al. (2012) Antibacterial effects of zinc oxide nanoparticles on Escherichia coli K88. Afr J Biotechnol 11: 10248-10254.
- 33. **Rajagopal R, Mini M (2013)** Outbreaks of salmonellosis in three different poultry farms of Kerala, India. Asian Pac J Trop Biomed 3: 496-500.
- 34. Akbar A, Anal KA (2013) Zinc oxide nanoparticles loaded active packaging, a challenge study against Salmonella typhimurium and Staphylococcus aureus in ready to-eat poultry meat. Food Control 38: 88-95.
- 35. Arabi F, Imandar M, Negahdary M, Imandar M, Noughabi MT, et al. (2012) Investigation of antibacterial effect of zinc oxide nanoparticles upon life of Listeria monocytogenes. Annals Bio Res 3: 3679-3685.
- 36. Gales AC, Jones RN, Turnidge J, Rennie R, Ramphal R (2011) Characterization of Pseudomonas aeruginosa isolates: occurence rates, antimicrobial susceptibilitypatterns and molecular typing in the global SENTRY antimicrobial surveillance program, 1997-1999. Clin Infect Dis 32: 146-155.
- 37. Hillier A, Alcorn JR, Cole LK, Kowalski JJ (2006) Pyoderma caused by Pseudomonas aeruginosa infection in dogs: 20 cases. Vet Dermatol 17: 432-439.

- 38. Li XZ, Livermore DM, Nikaido H (1994) Role of efflux pump(s) in intrinsic resistance of Pseudomonas aeruginosa: resistance to tetracycline, chloramphenicol, and norfloxacin. Antimicrob Agents Chemother 38: 1732-1741.
- 39. Nikaido H (1994) Prevention of drug access to bacterial targets: permeability barriers and active efflux. Science 264: 382-388.
- 40. *Hall-Stoodley L, Costerton JW, Stoodley P (2004)* Bacterial biofilms: from the natural environment to infectious diseases. Nat Rev Microbiol 2: 95-108.
- 41. Rosi NL, Mirkin CA (2005) Nanostructures in biodiagnostics. Chem Rev 105: 1547-1562.
- 42. Gunalan S, Sivaraj R, Rajendran V (2012) Green synthesized ZnO nanoparticles against bacterial and fungal pathogens. Progress in Natural Science: Materials International 22: 693-700.
- 43. Bosanquet AG, Bell PB (2004) Ex vivo therapeutic index by drug sensitivity assay using fresh human normal and tumor cells. J Exp Ther Oncol 4: 145-154.
- 44. Nie S, Xing Y, Kim GJ, Simons JW (2007) Nanotechnology applications in cancer. Annu Rev Biomed Eng 9: 257-288.
- 45. Alexis F, Basto P, Levy-Nissenbaum E, Radovic-Moreno AF, Zhang L, et al. (2008) HER-2-targeted nanoparticle-affibody bioconjugates for cancer therapy. ChemMedChem 3: 1839-1843.
- 46. *Mishra B, Patel BB, Tiwari S (2010) Colloidal nanocarriers: a review on formulation technology, types and applications toward targeted drug delivery. Nanomedicine 6: 9-24.*
- 47. Shapira A, Livney YD, Broxterman HJ, Assaraf YG (2011) Nanomedicine for targeted cancer therapy: towards the overcoming of drug resistance. Drug Resist Updat 14: 150-163.
- 48. Rasmussen JW, Martinez E, Louka P, Wingett DG (2010) Zinc oxide nanoparticles for selective destruction of tumor cells and potential for drug delivery applications. Expert Opin Drug Deliv 7: 1063-1077.
- 49. Gojova A, Guo B, Kota RS, Rutledge JC, Kennedy IM, et al. (2007) Induction of inflammation in vascular endothelial cells by metal oxide nanoparticles: effect of particle composition. Environ Health Perspect 115: 403-409.
- 50. Sayes CM, Reed KL, Warheit DB (2007) Assessing toxicity of fine and nanoparticles: comparing in vitro measurements to in vivo pulmonary toxicity profiles. Toxicol Sci 97: 163-180.
- 51. Beyerle A, Schulz H, Kissel T, Stoeger T (2009) Screening strategy to avoid toxicological hazards of inhaled nanoparticles for drug delivery: the use of alpha-quartz and nano zinc oxide particles as benchmark. Inhaled Particles 151: 1-9.
- 52. Lappin MB, Campbell JD (2000) The Th1-Th2 classification of cellular immune responses: concepts, current thinking and applications in haematological malignancy. Blood Rev 14: 228-239.
- 53. Hanley C, Layne J, Punnoose A, Reddy KM, Coombs I, et al. (2008) Preferential killing of cancer cells and activated human T cells using ZnO nanoparticles. Nanotechnology 19: 295103.
- 54. Dorfman A, Parajuli O, Kumar N, Hahm JI (2008) Novel telomeric repeat elongation assay performed on zinc oxide nanorod array supports. J NanosciNanotechnol 8: 410-415.

- 55. Shen W, Xiong H, Xu Y, Cai S, Lu H, et al. (2008) ZnO-poly(methyl methacrylate) nanobeads for enriching and desalting low-abundant proteins followed by directly MALDI-TOF MS analysis. Anal Chem 80: 6758-6763.
- 56. Valentine BA (2004) Neoplasia: In: Fubini SL, Ducharme NG. Farm Animal Surgery. Saunders.
- 57. Hassanein KMA, Mahmoud AZ (2009) Pathological studies on tumor incidence in farm animals. Alex J Vet Sci 28: 105-117.
- 58. Dobson JM, Samuel S, Milstein H, Rogers K, Wood JL (2002) Canine neoplasia in the UK: estimates of incidence rates from a population of insured dogs. J Small AnimPract 43: 240-246.
- 59. Vezzali E, Parodi AL, Marcato PS, Bettini G (2010) Histopathologic classification of 171 cases of canine and feline non-Hodgkin lymphoma according to the WHO. Vet Comp Oncol 8: 38-49.