Antimicrobial effected of red dragon(Hylocereus polyrhizus) on bacterial isolated from frozen meat.

Noor A. ALtaai

Unit of Zoonotic Diseases, College of Veterinary Medicine, University of Baghdad, Iraq

e.mail: noor.ham@covm.uobaghdad.edu.iq

Abstract:

The present study pointed to estimated the validation of red dragon fruit (Hylocereus polyrhizus) extracts against three microbial pathogens (*Staph. E.coli* and *Salmonella*)isolated from frozen meat samples.

One hundred twenty frozen meat samples were aseptically collected from local markets of Baghdad ,familiar bacteriological mechanism were used to isolate Staph. aureus, E. coli and Salmonella spp. Ecoli 35(29.1%) is high percentage, Staph .aureus 21(17.5%) and Salmonella 14(11.6%) isolates .The antibacterial efficiency of red dragon fruit (Hylocereus polyrhizus) extracts was determined by the agar well diffusion method to investigate the minimum inhibitory concentration (MIC) at different concentration started with 20, 1.25 mg/ml. The highest antibacterial activity between 20 10, 5, 2.5 and mg/mL and 5 mg/mL except in Salmonella spp. the value of inhibition was between 20 mg/mL and 10 mg/ml. The highest zone of inhibition of Staph. Auras (27 mm) While (22mm ,17mm)of E.coli and Salmonella respectively at 20 mg/mL. (21,22,12 mm) inhibition zone against Staph. aureus, E. coli and Salmonella spp. respectively at 10 mg/ml and (15, 10) mm against Staph. Aureus and E. coli at 5 mg/ml. Antimicrobial disc revealed all bacteria resistance to Bactircin, Amoxicillin clavulanic acid and Carbencillin disc, while sensitive to Tetracycline, and intermediate zone of inhibition to Ceftriaxon.

Key words: red dragon(Hylocereus polyrhizus), frozen meat ,antimicrobial, Staph, E.coli, Salmonella

Introduction:

Meat is renowned as a scope intermediate for pervasion food-borne diseases due to its high water action, high protein content, and approximately neutral pH, which create suitable statefor the growth of bacteria, contaminated meat depends on number ofbacteria, physicochemical properties, time/temperature of storageof meat (Mc Guinnesset al., 2009).

Generally, contamination occurs because of inappropriate salutary conditions and handling inslaughterhouses, the ability of bacteria across contamination by forming the biofilm, in addition to condition before slaughter as feeding and housing including contaminations from skin and feces by contents of digestion system (Doulgeraki *et al.*,2012).

Oneof the most common food poisoning is *Staphylococcal* and is of major concern in public health programsworldwide (Hennekinne *et al.*,2012). Food poisoning by *Staphylococcal* due to its produced enterotoxins, this enterotoxin characteristic by heating resistance (heat stable), acid-resistant, and resistant to proteolytic enzymes effect as pepsin and trypsin (Jablonski and Bohach 1997).

The normal flora in the intestine of humans and warm-blooded animals is *Escherichia coli*from Family Enterobacteriaceae causeshighly foodborne disease. Undercooked ground meat products, raw milk, and contaminated raw vegetables the most contamination in human (Castro *et al* .,2017)

Salmonellosis are major problem to causes Food borne diseases represent a for health in developed and developing countries, Over two thousand serotypes of *Salmonella*can cause food poisoning in worldwide as *S. typhimurium*, *S. enteritidis*, the most frequently isolated⁶. Risk of salmonellosis represented by the ingested bacteria and multiplying in the intestine of the host and invading the Gastro Intestinal tracts morethan to release toxins(Birmingham*et al.*, 2006).

Red Dragon fruit (Hylocereus polyrhizus) is a nutritious fruit cultivated throughout Asian countries particularly. The fruit have special shape and majestic color with mouthwatering taste in addition to have vital nutritional ingredients as carotene, calcium, fiber, vitamin B, vitamin C, and phosphorous (Kirti *et al.*,2020). Red Dragon fruit peel extract contain many the phytochemical compounds: Flavonoids, phenols, hydroquinones, and saponins. Also Steroids and triterpenoids composition found in peel therefore could stope the growth of bacteria (Ichsan *et*

al., 2018) .Red dragon fruit peel extract is also proved to be able to inhibit the growth of Salmonella pullorum and Staphylococcus aureus(Arief et al.,2015).

The current investigation was conducted to estimate the bacterial load of Meat in local Iraqi markets and estimated antimicrobial activities of the red dragon fruits extracts were associated with high phytochemical compounds and total phenols contained in the extracts.

Material and methods:

Area of study:

The study was conducted in Baghdad Province, Iraq. was done in the Bacteriology Laboratory, Department of zoonosis unite, Faculty of Veterinary Medicine, University of Baghdad.

Frozen Meat sample collection:

One hundred twenty frozen meatsamples were collected from local markets in Baghdad city ,the samples were packed, identified,transferred in ice-box and immediatelyprocessed at zoonosis laboratory –veterinary medicine collage.

Preparation of samples:

The samples were cut with a sterile knife, 25 g of each samples were added to 225ml buffered peptone water 0.1% (Bakhtiary *et al* .,2016). The samples were then homogenized for 2 minutes. Ten fold serial dilutions were prepared for further analysis.

Staphylococcus isolation:

One ml of sample suspension added to Mannitol salt platesagar then incubated at 37°C for 24 hours. Gram stain and Biochemical tests was done to identified *S. auras* as catalase activity and coagulase tests.(Bannerman, 2003). Gram stain, catalase enzyme, coagulase and other common biochemical tests were used to confirm the identity of S. aureus.

E.coli isolation:

One ml of samples suspension added to MaCconky plates agar and Eosin methylene blue agar incubated at 37°C for 24 hours. Gram stainand Biochemical tests was done to identified *E.coli* by using The API-20E test kit for the identification of enteric bacteria(MacFaddin ,2000).

Salmonella isolation:

One ml of samples suspension added to 10 ml Tetrathionate broth incubated at 37°C for 24 hours then inoculated the enrichment broth on Xylose Lysine Desoxycholate (XLD) agar and Salmonella-Shigella (S.S) agar incubated at 37°C for 24 hours. Gram stain and Biochemical tests was done to identified *Salmonella* by using The API-20E test kit for the identification of enteric bacteria (Koneman *et al.*,2005).

Preparation of red dragon fruit extract:

Dragon fruit was obtained from the popular Malaysia market ,kept cold . then washing with cleaned water and cut the red dragon fruit into small pieces (2mm), dried for 4 days , use the blender until its became powder then weighed .

Soak the dry fruit in 96% ethanol Sterilization as ratio 7.5 times the weight of the powder, filter the maceration by using filter paper. Then dry it by using a rotary evaporator at 45-50 °C.kept the extract at 4°C until further uses. making extract concentration of (200, 150, 100 50, and 25 mg/ml) (Rahayu *et al.*, 2019).

Preparation of microorganism and inoculums:

organisms were culturing on brain heart infusion agar overnight, then used for the preparation of bacterial suspensions diluting overnight cultures in saline and adjusted to 0.5 McFarland turbidity standards to approximately 108 CFU ml for each bacterium.

Agar well diffusion method for antibacterial activity:

Mueller Hinton agar (MHA, Oxoid), media were prepared according to the instructions of manufacturers company. The Petri plates cultures by bacterial tested inoculated (0.2 ml each) using the sterilized swabs then a sterilized stainless steel borer was used to formed five wells (6 mm diameter), the red dargon extract was dissolved in sterile distilled water at concentrations of (20, 10, 5, 2.5 and 1.25 mg/ml), then 100 µl of each concentration of the plant extract were filled each well. The Petri plates were incubated at 37°c for 24 hours in an incubator and zone of inhibition(mm) was observed and measured using a scale around the extracts (Lino and Deogracios, 2006).

Antimicrobial Susceptibility Testing:

The teste done according to the Kirby-Bauer method by diffusion of antibiotics on the surface of Muller-Hinton agar. the preparation of bacterial suspensions diluting overnight cultures in saline and adjusted to 0.5 McFarland turbidity standards to approximately 108 CFU ml for each bacterium then inoculated Muller-Hinton agar with bacterial suspension by sterile swab (0.2 ml each) with dry the plates for ten minutes .Put the disk with 15 mm from the edge, measured the inhibition zone using a scale around the antibiotic disc(Bauer *et al.*,1966).

Result and discussion:

According to table(1), figure (1) the percentage of contaminated Staph. Aureus Salmonella and E.coli in frozen meat in current study show that the isolation)is high percentage, staph 21(17.5% rates of *Ecoli* 35(29.1%) and Salmonella 11.6%) isolates conferring to their morphological characterization and biochemical tests, this results agreement with (Abdelraouf et al., 2013) who found 11 (7.3%) samples were positive with Salmonella in Gaza strip ,also(Eman et al .,2014) found Escherichia coli (40%) and Staphylococcus aureus (29%) in butchers shops, with E.coli (19%) ,S.aureus (28%) and Klebsiella sp. (9%) in restaurants, while Other reports recorded that S.aureus frequently are present in low numbers on raw meat surface occurs infrequently (Saadia and. Hassanein(2010). The reason of contamination may be to the location of the animal (housing) ,feeding in addition to and the instrument used in slaughter the animals and places of slaughter as well as the butchery itself and the contamination of hands and even the wood used to cut flesh and without sterilization

Table (1): percentage of *Staph. Aureus, Salmonella* and *E.coli*isolated from frozen meat:

samples	Bacterial isolated	Gram stain	Percentage
Frozen Meat (120	Staph .auras	G+	21(17.5%)
)samples	E.coli	G-	35 (29.1%)
	Salmonella	G-	14(11.6%)

40 35 35 No. of samples (120) 30 25 21 20 14 15 10 5 G+ G-G-E. coli salmonella staph. Auras Axis Title

Figure (1):percentage of *Staph. Aureus*, *Salmonella* and *E.coli* isolated from frozen meat

Morphology and biochemical test:

All staphylococcal isolates from different samples showed the colonies of *Staphylococcus aureus* on the Mannitol salts agar utilization of salt and give yellow color, coagulase positive, oxidase negative, catalase positive, and DNase positive, in addition beta hemolysis test on blood agar (Table: 2) hemolysis (α , γ or β hemolysis) on blood

Table (2): Result of Biochemical reaction of *S.aureus***:**

Biochemical reaction of S. aureus	
Slide Coagulase test	+
tube Coagulase test	+
Oxidase test	-
Catalase test	+

All *E.coli* isolates from different samples fermentation the lactose in MaCconky agar and give pink in color and give metallic sheen in Eosin methylene blue agar with results of API-20E test kit for the identification of enteric bacteria.

Salmonella isolates give pink colonies with black centers on Xylose Lysine Desoxycholate (XLD) agar, and pale, smooth, rounded black centered colonies on

Salmonella-Shigella (S.S) agar and results of API-20E test kit for the identification of enteric bacteria.

Antimicrobial effect of red dragon fruits:

In our study and according to table (3) the inhibition zone of red dragon fruit (Hylocereuspolyrhizus) extract against the growth of *Staph. Aureus*, *Salmonella* and *E.coli* show the biggest drag zone found against *staph .aureus* (27 mm), (22 mm) against *E.coli* and (17mm) against *Salmonella* in 20mg/ml.

while S. aureus, E.coli and Salmonella had 21mm,16 and 12mm respectively in 10 mg/ml of the extract concentration .

At 5 mg/ml showed the highest zone of inhibition *in S.aureus*15mm and *E.coli* 10mm while in *Salmonella*not sensitive to the extract all bacteria were not sensitive to the extract at 2.5mg/ml and 1.25 mg/ml. (fig.2).

Our results revealed the highest zone of inhibition Gram-positive bacteria, *S. aureus*, was extra susceptible to the antibacterial activity of the red dragon fruit peel extract are agreement with(Manihuruka *et al.*,2017) who reported High inhibition zone produced by red dragon fruit against *staph. aureus* and supported by Arief *et al.*, 2015)who analysis that the Gram-positive bacteria were more susceptible to antibacterial activity due to the absence of a lipoprotein wall that capable of preventing antimicrobial compounds.

The inhibition zone observed in gram negative bacteria *E.coli* with medium zone and less in *Salmonella* according to results are conducted by (Nurmahani *et al.*, 2012). The inhibit the growth occur due to Phenolic compounds in the extracts of pomegranate peel Phenolic compounds may inhibit the growth of bacteria with the ability of damaging the cytoplasmic membrane and proteins as well as inactivated some bacterial enzymes (Choi*et al.*,2011).

Antibacterial activity against both bacteria was also observed by (Tahera et al .,2014).

Table (3): The average of inhibition zones of red dragon fruit against *Staph*. *Aureus, Salmonella* and *E.coli*:

Concertation of	Zone of inhibition (mm diameter)			
red dragon	Staph. Auras	Salmonella	E.coli	
1.25mg/ml	-	-	-	
2.5 mg/ml	-	-	-	
5mg/ml	15	-	10	
10mg/ml	21	12	16	
20mg/ml	27	17	22	

Figure (2): The average of inhibition zones of red dragon fruit against Staph. Aureus, Salmonella and E.coli:

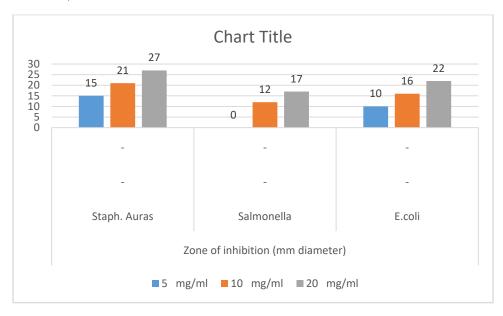


Table (4) revealed different results of microbial against the antimicrobial disc. All bacteria showed resistance to Bactircin, Amoxicillin clavulanic acid and Carbencillin disc against the antimicrobial disc .The study showed that all the isolates were sensitive to Tetracycline, also all isolated intermediate zone of inhibition to Ceftriaxon .

Our results matching with (Eman et al .,2014) who showed S.aureus isolated from fast food samples more resistance rate to commonly used (Lincomycin ,

Erythromycin and) and E. coli low resistance to amoxicillin. Also agreement with (Cook*et al* .,2009) reported about the antimicrobial resistance in *Campylobacter*, *Salmonella*, and *Escherichia coli* isolated meat from southern Ontario.

In Saudi Arabia (Al-Humam, 2019)showed Antimicrobial susceptibility of 40% of E. coli isolates to be ESBL is higher than (Bouchillon *et al* .,2004) who reported Enterobacteriaceae, vancomycin-resistant *Enterococcus faecium* and methicillin-resistant *Staphylococcus aureus* from some European countries.

Table (4): Mean of inhibition zone diameters of antibiotics:

Antibioti	Ceftriax	Trimeth	Genta	Doxyc	Azithro	Carben	Tetracy	Amoxic	Bacti
c	on	eprime	mycin	yclin	mycine	cillin	cline	illin	rcin(
	(CRO)	(TMP))	(CN)		(AZM)	(Py))	(TC)	clavulan	B)
								ic acid	
Staph	I	I	S	S	S	R	S	R	R
E.coli	Ι	R	R	I	I	R	S	R	R
Salmonell	Ι	S	S	I	S	R	S	R	R
a									

R:resistance, I: intermediated, S:sensitive

Conclusion:

Based on the results of this study its concluded the red dragon fruit extract (Hylocereuspolyrhizus) have antibacterial activity against *Staph. aureus E.coli* and *Salmonella*. The red dragon fruit extract (Hylocereuspolyrhizus) conceder Natural antibacterial material.

References:

Abdelraouf A, Elmanama A, Kamal A E, El kahlout, Shorok R, Elnakhala(2013). Occurrence of Salmonella in Fresh and Frozen Meat in the Gaza Strip Journal of Purity, Utility Reaction and Environment . 6(2) 142-152.

Al-Humam (2019).

NADetection of Escherichia coli , Salmonella spp. and Staphylococcus aureus in Ready-to-Eat Food in Al-Ahsa Province, Saudi Arabia .J Nutr Food Sci.; 9 (2); No: 754.

Arief II, Budiman C, Jenie BSL, Andreas E, Yuneni A (2015). Plantaricin IIA-1A5 from Lactobacillus plantarum IIA-1A5 displays bactericidal activity against Staphylococcus aureus. Beneficial Microbes. 6: 603–613.

Bakhtiary F, Hamid R S, Marlene R, Berit H, Hedayat H, Alexander G. H(2016). Evaluation Of Bacterial Contamination Sources In Meat Production Line. J. Food Quality.; 39: 750–756.

Bannerman TL(2003). Staphylococcus, Micrococcus, and other catalase positive cocci that grow aerobically. In: Murray PR, Baron EJ, Jorgensen JH, et al. Editors. Manual of clinical microbiology. Washington, DC: ASM Press. 2003;33;384-404.

Bauer A W, Kirby WMM, Sherris JC, Turck M(1966) .Antibiotic susceptibility testing by a standardize single disk method. Am. J. Clin. Pathol .45:493-496.

Birmingham CL, Smith AC, Bakowski MA (2006). Autophagy controls Salmonella infection in response to damage to the Salmonella-containing vacuole. J Biol Chem . 281(16):11374–11383.

Bouchillon SK, Johnson BM, Hoban DJ, Johnson JL, Dowzicky MJ, Wu DH,(2004). Determining incidence of extended spectrum β-lactamase producing Enterobacteriaceae, vancomycin-resistant Enterococcus faecium and methicillin-resistant Staphylococcus aureus in 38 centres from 17 countries: The pearls study 2001-2002. Int J Antimicrob Agents. 24(2):119-124.

Broz P, Ruby T, Belhocine K (2012). Caspase-11 increases susceptibility to Salmonella infection in the absence of caspase-1. Nature. 2012; 490(7419):288–291.

Castro VS, Carvalho RCT, Conte-Junior CA, Figueiredo EES (2017). Shigatoxin producing Escherichia coli: pathogenicity, supershedding, diagnostic methods, occurrence, and foodborne outbreaks. Compr Rev Food Sci Food Saf.;16(6):1269-1280.

Choi, J G, Kang O H, LeeY S, Chae, HS. Oh, Y C, Brice O O, Kim M S, Sohn D H, Kim H S, Park H, Shin D W, Rho J R, Kwon D Y. (2011). In vitro and in vivo antibacterial activity of punica granatum peel ethanol extract against Salmonella. Evid. Based Comp. Alternat. Med. 1-8. https://doi.org/10.1093/ecam/nen085

Cook, A, Reid-Smith, R, Irwin R, McEwen SA, Valdivieso-Garcia A. Ribble C. (2009). Antimicrobial resistance in Campylobacter, Salmonella, and

Escherichia coli isolated from retail turkey meat from southern Ontario, Canada.J. Food Prot. 72: 473-481.

Doulgeraki AI, Ercolini D, Villani F, Nychas GJE (2012). Spoilage Microbiota Associated To The Storage Of Raw Meat In Different Conditions. Int. J. Food Microbiol.;157: 130–141.

Eman M. Jarallah , Seaab I S, Khalid Y.(2014) . Isolation and Identification of some pathogenic Bacterial Species Contaminated from Meats in Butchers Shops and Kebab Restaurants in AL-Kut city Euphrates Journal of Agriculture Science-6.; (4): 30-37 .

Hennekinne JA, De Buyser ML, Dragacci S (2012). Staphylococcus aureus and its food poisoning toxins: characterization and outbreak investigation. FEMS Microbiol Rev. 36 (4): 815-836.

Ichsan MI, Abdul HA, Fakhrurrazi, Dewi M, Jalaluddin M (2018). Antibacterial test of red dragon fruit extract peel (hylocereus polyrhizus) against bacteria salmonella pullorum. J Med Vet . 12:9-14.

Jablonski LM, Bohach GA (1997). Staphylococcus aureus. food microbiology fundamentals and frontiers. (Doyle MP Beuchat LR & Montville TJ Edn). Washington, DC: ASM Press:353-357.

Kirti J M, Kumar M, Bhushan B, Pankaj K (2020). Postharvest Profile, Processing and Waste Utilization of Dragon Fruit (Hylocereus Spp.): A Review, Food Reviews International.; 8755-9129: 1525-6103.

Koneman WK, Allen SD, Janda WM, Schreckenberger PC, Propcop GW, Color Atlas and Textbook of Diagnostic Microbiology, 7th Edn. Lippincott-Raven Publisher, Philadelphia, USA.(2005)...

Lino, A.and O. Deogracios (2006). The in-vitro antibacterial activity of Annona senegalensis, Securidacca longipendiculata and Steanotaenia araliacea-Ugandan Medicinl plants. Afri. Health Sci .6(1): 31-35.

MacFaddin, J.F.(2000) Biochemical tests for identification medical bacteria. Warery press, INC. Baltimore, Md. 21202 USA.

Manihuruka F M Suryatib, T, Ariefb I I (2017). Effectiveness of the Red Dragon Fruit (Hylocereus polyrhizus) Peel Extract as the Colorant, Antioxidant, and Antimicrobial on Beef Sausage. Media Peternakan, 40(1):47-54

Mc Guinness S. H, McCabe E, O'Regan E, Dolan A, DuffyG,(2009).action of a rapid real-time PCR based method for the specific detection of Salmonella on fresh meat .Meat Science. ;83: 555–562.

Nurmahani M M, Osman A, Hamid A A, Ghazali FM, Dek M P(2012). Antibacterial property of Hylocereus polyrhizus and Hylocereus undatus peel extracts. Int. Food Res. J.;19: 77-84.

Rahayu Y C, Ardo S, Dyah S (2019). Antibacterial Activity of Red Dragon Fruit Extract (Hylocereuspolyrhizus) On Streptococcusmutans International journal of applied pharmcutics. 11(4);0975-705.

Saadia, M. Hassanein, E.(2010).The Microbial Quality of Fast Food and Traditional Fast Food. Nature and Science. (10):8.

Tahera J,Feroz F, Senjuti J D, Das K K, Noor R. (2014). Demonstration of anti-bacterial activity of commonly available fruit extracts in Dhaka, Bangladesh. American J. Microb. Res. 2: 68-73. https://doi.org/10.12691/ajmr-2-2-5