Isolation and Identification of Indole Acetic Acid (IAA) Producing Bacteria from Organic Soil: Investigating Its Efficacy on Plant Growth

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ABSTRACT:

Indole acetic acid (IAA) production is a major property of rhizosphere bacteria that stimulate and facilitate plant growth. The present work deals with isolation, characterization and identification of indole acetic acid producing bacteria from the rhizospheric soil. Out of four organisms isolated from serial dilution, Sample-2 confirmed thick bands of IAA production by TLC analysis. 16s rRNA analysis showed the isolated organism was found to be *Pseudomonas fluorescence* with similarity of 88%. The isolated organism treat plants showed increased shoot and root length in plant growth analysis.In conclusion the isolated IAA producing bacteria can be used as efficient biofertilizer inoculants to promote plant growth.

KEYWORDS: Tryptophan, rhizobacteria, purification, optimization, plant nutrition

INTRODUCTION:

Biofertilizers are one of the excellent tool for sustainable agriculture. Biofertilizers improves soil fertility through solving atmospheric nitrogen, each, in affiliation with plant roots and with out it, solubilize insoluble soil phosphates minerals and produces plant boom materials with inside the soil.Biofertilizer incorporates microorganisms which provides good enough deliver of vitamins to the host flora and make sure their right improvement of growth and law of their physiology. Effective dwelling microorganisms are used in the instruction of biofertilizers. (Sadaf Shahab et al,2009).

The common production of plant growth regulators are (auxin, gibberellin,ethylene etc.), siderophores, HCN and antibiotics (Arshad and Frankenberger, 1992). Indole acetic acid is an crucial (IAA)physiologically lively auxins. IAA is commonly made from L-tryptophan metabolism via way of means of numerous microorganisms consisting of PGPR(Shahab S et al,2008). Microorganisms inhabiting rhizospheres of numerous plant life are in all likelihood to synthesize and launch auxin as secondary metabolites due to the rich elements of substrates exuded from the roots as compared with non rhizospheric soils (Kampert et al., 1975). Plant growth promoting rhizobacteria (PGPR) is a micro organism that actively colonize plant roots and has a more yield and greater growth.

One of the alternative maximum crucial powerful elements in growing plant yield is seed inoculation or priming with plant growth promoting rhizobacteria (PGPR). Also, plant growth promoting rhizobacteria (PGPR) are a collection of micro organism that actively colonize plant roots and growth plant increase and yield (Subba RaO 1999; Wu et al., 2005; Heidari et al.,2011). Some chemical fertilizers in use incorporate poisonous through-additives inclusive of heavy metals, inorganic acids and natural pollutants, and hence, a long-time period utility of those chemical fertilizers can probable set off the buildup of those with the aid of using-additives in soil, leading to the worsening of soil ecological environment, and making the heavy metals, nitrate, and different dangerous additives in agricultural merchandise consisting of greens, grains and culmination critically passed the standards(Verma J.P et al., (2014)).

Increasing agricultural productiveness is presently related to the sizeable software of chemical fertilizers. However, issues like better cost, pollutants of herbal assets and fitness implications have forced researches to discover options for secure and expanded crop productiveness. One such opportunity is using PGPB, especially IAA generating microorganism.Beneficial microbial allelopathies in rhizosphere are a key agent of alternate in soil environment and have an effect on crop fitness and yield (Sturz and Christie, 2003). Due to such precious attribute, isolation of indigenous bacterial lines with plant boom selling capacity from diverse environments stays a famous concept. In this regard, the existing observe turned into performed to isolate capacity IAA producing micro organism.

Indole acetic acid (IAA) directs numerous factors of plant growth and development, including the induction and regulation of a variety of processes: e.g., Cellular department, root extension, vascularization, apical dominance, and tropisms. The outcomes of IAA on plant root tissue are attention based and may be species specific. Responses to growing IAA concentrations strengthen from the stimulation of number one root tissue to the improvement of lateral and adventitious roots and eventually to the entire cessation of root increase (Patten, C. L., and B. R. Glick. 2002).

Indole acetic acid (IAA) is thought to stimulate each speedy responses (e.g. will increase in cellular elongation) and longtime responses (e.g.cellular department and differentiation) in flowers. L-tryptophan (L-Trp), is an amino acid, serves as a physiological precursor for biosynthesis of IAA in flowers and in microbes. Root exudates are herbal supply of L-Trp for rhizospheric microflora, which might also additionally decorate IAA biosynthesis in the rhizosphere. Microbial biosynthesis of IAA in soil is greater through L-Trp from root exudates or decaying cells. The reaction of cereals and greens to inoculation with a few PGPR micro organism has been attributed to the impact of plant growth regulators launched through these microorganisms (Flashman MA et al., 1996).

The present study concentrates on identity of IAA producing microorganism from agricultural land. The screening was done through TLC method. The highest IAA producer is identified by 16s rRNA sequencing technique. Plant growth analysis was carried out to determine the efficacy of the IAA producer.

MATERIALS AND METHODS:

Sample series and isolation of Bacteria:

Place serially diluted soil samples (0.5 ml) on solidified nutrient agar medium, even as the use of spread plate and pour plate strategies to domesticate organisms from the samples. After all of the liquid in the medium has been absorbed, the petri dish is sealed and incubated in the dark at 37°C for twenty-four hours. Select colonies of various shapes and switch to nutrient agar medium. After numerous subcultures, natural isolates have been sooner or later received. Isolates examined in the microscope have been preserved in nutrient broth (NB) medium containing 15% glycerol at -20°C.

Production of IAA and TLC Analysis:

The decided on adverse bacterial lines had been cultured in 5ml nutrient broth medium in a test tube for24 hours. TLC slides have been organized with silica gel G and calcium carbonate.Propanol:Water (8:2) is used in the solvent system. Spot the extracted pattern and fashionable IAA (10mg/100ml) at the TLC plate.

DNA Isolation from Sample (Kit method):

Add 750 μ l of homogeneous 1X Buffer to the tradition pellet, vortex and maintain in a 65°C dry bath for 20 mins, then upload 750 μ l of Lysis buffer and blend thoroughly. Then keep it in the dry water bath at 65° C and centrifuge at 12,000 rpm for 10 mins Transfer the supernatant to a 2ml check tube, upload an identical extent of isopropanol and blend it manually, switch to the column(750 μ l every time) and spin at 12000 rpm for 1 minute, upload 750 μ l wash buffer, then spin at 12000 rpm for 1 minute. Repeat the bathing buffer step, dry spin for two mins, upload 50 μ l of elution buffer, and spin at 12000 rpm for 1 minute. The DNA consequently acquired ought to be quantified.

DNA quantification:

Estimate the DNA concentration by the means of recording the absorbance at 260 and 280 nm in a UV/VIS spectrophotometer. Collect 10 μ l of genomic DNA pattern in a quartz cuvette. Make a extent of 2ml with distilled water, measure the absorbance at 260 and 280 nm by the use of a UV spectrophotometer, calculate the ratio of A260/A280, and use the connection of

double-stranded DNA (O.D) to calculate the DNA attention. At 260 nm = 50 g/ml PCR situations: Initial denaturation at 95°C for two mins; very last denaturation at 95°C for 30 seconds; annealing at 55°C for 30 seconds; elongation at 72°C for 1 minute; repeat steps 2, 3 and 4 for 30 cycles. The last elongation is for 10 minutes at 72°C ant it is permanently stored in refrigerator at 4°C.

Preparation of PCR reaction mixture:

Each PCR response used amplification performance of DNA barcode primers and carry out multiplex PCR evaluation consists of 1µ1 DNA template (25 ng), 2µ1 10X response buffer, 0.5µ1 MgCl2 (50pM), 1µ1 dNTPs blend (10mM), 1µ1 ahead primer (10pM), 1µ1 opposite primer (10pM), 0.5µ1 Taq polymerase (five U/pi) volume 25µ1 can be adjusted with molecular grade water. Primers are fashionable primers to be had for 16srRNA gene amplification. Gel purification protocol Cut the preferred DNA band at the gel, upload 600µL of gel dissolution buffer 55°C till the gel is absolutely dissolved. Add 200µL of isopropanol, blend and switch to the chromatographic column, spin 6at 12000 rpm for 1 minute, upload 700 µL washing buffer, spin at 12000 rpm for 1 minute.

Sanger Sequencing PCR:

Initial denaturation at 95°C for 2 mins; very last denaturation at 95°C for 30 seconds; annealing at 50°C for 30 seconds; termination at 60°C for 4 mins; repeat steps 2, 3 and 4 for 30 cycles. Always preserve it at 4°C.

Post Sequencing and PCR Purification:

Add a 125mm 2.5 μ L EDTA to every properly and spin briefly. Use a multichannel pipette to upload 35 μ L of ethanol.Mix for 10 minutes at 2000 rpm and centrifuge for 30 minutes at 3510 rpm.Use a tissue bed to decanter the ethanol at 300rpm (flip the plate over for 30 seconds).Add 40.Repeat the above referred to invert spin.Air dry for 30-45mins overlaying the plate with lint loose tissue.Add 13 μ L of HiDi Formamide short spin .Denature at 95°C for 5 mins.Place the plate in sequencer.

Data analysis:

The received sequencing record might be in AB1 format, viewed through FinchTV, BioEdit, ChromasLite, SeqScanner and different software. The quality of the received series may be discovered via the peaks of the electropherogram. Use BLAST server or a server associated with a particular database to research sequencing data.

Plant growth analysis:

Take 4 pots of same length and quantity. Fill the pot with sterile soil and moisten it with sterile distilled water. *Spinacia oleracea* (pasalai keerai) seeds have been taken and sown in pots. Before sowing, sterilize the seeds with the running tap water, after which positioned them in a full of distilled water. A pinch of SDS was added and seeds have been shaken lightly for 1–2 min. The water was discarded and 70% ethanol was added for 1–2 min. Ethanol become discarded and seeds have been positioned in the sterile distilled water, a pinch of HgCl2 was added to this and kept undisturbed for 15 minutes.Using distilled water, it is washed for another 3-4 times, all the water was discarded. After the surface is sterilized, the seeds are inoculated. The pots are marked according to way they are handled. The seedlings was gently removed after 10 days from the soil immersing the seedling in water to get rid of the soil. Evaluate the dry weight of the plant, shoot length and root length(cm).

Shoot length and root length:

Three plantlets have been randomly decided on to report the basis duration and shoot duration of the *Spinacia oleracea* (pasalai keerai) plant. They are measured through the use of a centimeter scale. Fresh weight (mg/g dry wt.) of three plant samples had been randomly decided with the experimental pot. They are divided into roots and shoots. Their dry weight is measured by means of the use of an electronic single scale.

RESULTS:

Collection of soil and serial dilution:

Soil samples have been accumulated from agricultural land, Coimbatore. The place surrounding the roots of the plant was collected and saved in a container. Figure-1 indicates the sample collected place. Further the samples had been on nutrient agar. Different dilutions have been taken and plated. Figure-2shows the dilution from 10^{-2} to 10^{-7} .

TLC analysis and Fluorescence production analysis:

Thin layer chromatography evaluation was done to identify the IAA producing microorganism. Four bacteria have been isolated from serial dilution which confirmed predominant growth on plates. All the 4 bacterial metabolites had been screened for IAA production. Sample-2 confirmed thick bands which verify the better IAA production (Figure-3). Therefore, bacterial sample-2 is chosen for further evaluation. From the fluorescence production, when seen under UV light the identified strain showed positive result(Figure-4).

PCR amplification and species identification:

From PCR amplification, the amplified product observed to be 850bp. Figure-5 indicates the bands of agarose gel electrophoresis. Further the amplified 16s rRNA gene was subjected for sequencing. The identified sequence was shown below and the chromatogram was shown in Figure-6.

Coding sequence:

AAGTTGACCGCCTGGGGAGTACGGCCGCAAGGTTAAAACTCAAATGAATTGACGGG GGCCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAAGAGCCTTA CCAGGCCTTGACATCCAATGAACTTTCTAGAGATAGATTGGTGCCTTCGGGAACATT GAGACAGGTGCTGCATGGCTGTCGTCAGCTCGTGTGGGGAGATGTTGGGTTAAGTCC CGTAACGAGCGCAACCCTTGTCCTTAGTTACCAGCACGTAATGGTGGGCACTCTAAG GAGACTGCCGGTGACAAACCGGAGGAAGGTGGGGATGACGTCAAGTCATCATGGCC CTTACGGCCTGGGCTACACACGTGCTACAATGGTCGGTACAGAGGGTTGCCAAGCC GCGAGGTGGAGCTAATCCCTCAAAACCGATCGTAGTCCGGATCGTAGTCTGCAACTC GACTGCGTGAAGTCGGAATCGCTAGTAATCGCGAATCAGAATGTCGCGGTGAATAC GTTCCCGGGCCTTGTACACACGCCCGTCACACCATGGGAGTG GGTTGCACCAGAAGTAGC

BLAST analysis:

The similarity was determied among standard sequence and DNA base sequence with the help of BLAST method and the Query cover was displayed which shows the similarity percent of approximately 88% with *Pseudomonas fluorescens*. Since the similarity percent is low, high risk for the new strain.Figure-7and Figure-8 suggests the BLAST outcomes and BLAST graphic summary result.

Pot assay:

The Figure-9 shows plants with the aid of using pot assay approach. The seeds of *Spinacia oleracea*(pasalai keerai) have been grown on three exclusive pots with one pot which suggests the control, the any other pot handled with crude and the opposite with partly purified IAA. The table indicates the effects of shoot length, root length and fresh weight of the vegetation. From the Table-2, it shows the outcomes received, the plant handled with the partially purified IAA suggests the more growth in all factors, than evaluating with crude and water.

DISCUSSION:

Indole acetic acid (IAA) manufacturing is a major property of soil micro organism specifically Pseudomonas and Bacillus. The present work deals with indole acetic acid generating micro organism from the soil of various district of Uttar Pradesh. Optimization of indole acetic acid production was done at unique cultural conditions of pH and temperature with various media additives including carbon and nitrogen supply, tryptophan attention (Shih-Yung ,2010).

Indole-3-acetic acid (IAA) is the maximum studied auxin in plant life, and its biosynthesis pathway has been investigated for over 70 years. Although the entire picture of auxin biosynthesis stays to be elucidated, remarkable development has been made these days in umderstanding the mechanism of IAA biosynthesis. Genetic and biochemical research show that IAA is specially synthesized from L-tryptophan (Trp) through indole-3-pyruvate by means of two-step reactions in Arabidopsis. While IAA is likewise made from Trp through indole-3acetaldoxime in Arabidopsis, this pathway likely performs an auxiliary position in plant life of the own circle of relatives Brassicaceae. Recent research recommend that the Trp-independent pathway isn't always a chief path for IAA biosynthesis, however they monitor an essential position for a cytosolic indole synthase on this pathway (Pollmann, Neu, Weiler,2003).

IAA generating microorganism from rhizosphere soil and their impact on seed germination changed into investigated by means of Sharnali Das *et al.* Out of 30 isolates, eight (viz. Aux four, Aux 9, Aux14, Aux16, Aux 19, Aux 20, Aux 21 and Aux 25) have been able to produce IAA which was confirmed by Salkowski reagent test. Isolates had been characterised on the basis of visible observations, gram staining and bio chemical assessments together with oxidase, catalase, indole, TSI, methyl red, Voges-Proskauer, citrate utilization, urease and starch hydrolysis test. The IAA producing bacteria are rod and spherical formed gram Positive and gram negatIvemicroorganism. IAA has been implicated in really all elements of plant growth and improvement inclusive of the developing of longer roots with elevated variety of root hairs and root laterals that are concerned in the nutrient uptake (Datta and Basu, 2000). Furthermore, it complements embibal activity, inhibit or delay abscission of leaves and promote flowering and fruiting in plant (Zhao, 2010).

This study suggests that the *Spinacia oleracea* (pasalai keerai) seeds have been sown in 3 pots (pot assay). The seeds that have been dealt with partially purified IAA confirmed larger growth of shoot length, root length and fresh weight than the seeds dealt with crude and water (control). Dobbelaere *etal* (2003) and Cakmakı (2005a) were reported that PGPR can increase yield and leaf area index, shoot and root weight and delay leaf senescence. Numerous different research have proven a huge increase in dry matter accumulation and seed yield while inoculated

with PGPR (Perveen et al., 2002; Waniet al., 2007; Mishra et al., 2010; Sharifiet al., 2011). Dilfuza (2007) stated that once corn seeds are inoculated with Azospirillum brazilance indicates elevated dry matter accumulation. Mishra et al., (2010) said that the maximum of diagnosed isolates shows significant increase of each shoot and root of *Cicerarietinum* seedlings.

The outcomes of the study by Sharifiet al., (2011) confirmed that the seeds inoculating with Plant Growth Promoting Rhizobacteria confirmed grain yield, plant height, quantity of kernel in keeping with ear, variety of grains according to ear row significantly. Strains of *Pseudomonas putida* and *Pseudomonas fluorescens* increases root and shoot elongation in canola (Glick et al., 1997).Pseudomonas and Bacillus micro organism posses numerous plant growth promoting activity. Compare to those microorganism, *Pseudomonas fluorescence* have ability to supply excessive quantity of IAA manufacturing than *Bacillus subtilis* and control. The end result shows that those isolates consists of PGP activities, that percent of IAA manufacturing of those bacterial activities are numerous greatly. Mahalakshmi and Reetha (2009), (Saitou N and Nei M 1987) said identical consequences in checking out PGP activities of bacteria which are collected from the rhizosphere of tomato.

The rhizosphere microorganism specifically *Pseudomonas fluorescens*, have extra capacity to grow the host plant through numerous mechanism to decrease the plant illnesses which incorporates manufacturing of effective siderophores (O sullivan and O Gara 1992): Has and Defago 2005). These like*Pseudomonas fluorescens* and different microorganism Bacillus *subtilis* had been in addition studied or their impact on plant growth under controlled situations.Data received from the pot experiments established tremendous effects on root elongation of handled plant life over the control. This shows that those micro organism have the performance to enhance the size of roots and shoots of the plant and those may be taken into consideration as plant growth promoters.

CONCLUSION:

Out of four organisms isolated from serial dilution, Sample-2 confirmed thick bands of IAA production by TLC analysis. 16s rRNA analysis showed the isolated organism was found to be *Pseudomonas fluorescence* with similarity of 88%. The isolated organism treat plants showed increased shoot and root length in plant growth analysis. In conclusion the isolated IAA producing bacteria can be used as efficient biofertilizer inoculants to promote plant growth.

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Sl.No	COMPONENTS	CONCENTRATIONS
1	Forward primer (27F)	10pM
2	Reverse primer (1492R)	10pM
3	10Mm dNTPs	10µM
4	10X PCR Buffer	2x
5	25Mm Mgcl ₂	2.5mM
6	Template DNA	25-40ng
7	Taq polymerase	5U
8	Nuclease free water	Make up to 25 µl

Table 1: PCR reaction mixture

Table 2: Pot assay

S.NO	Shoot length (mm)	Root length (mm)	Fresh weight (g)
Control	8	10	0.030
Crude	12	11	0.045
Partial purified	22	18	0.08



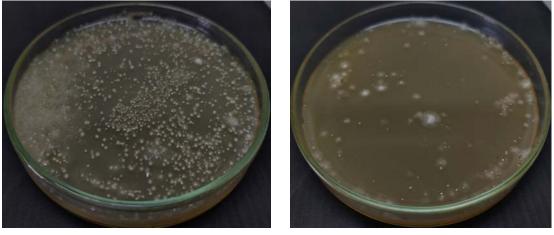
Figure-1: Collection of soil sample



10⁻² dilution



10⁻⁴ dilution



10⁻⁵ dilution 10⁻⁷ dilution Figure-2:Serial dilution of soil samples

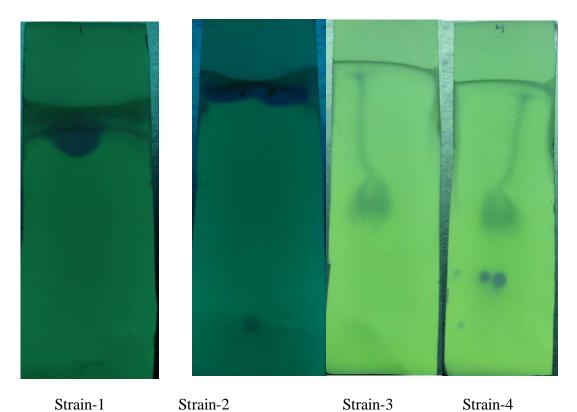
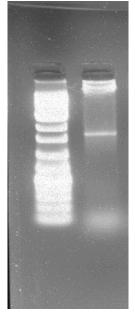


Figure-3: Thin layer chromatographic analysis of isolated bacteria



Figure-4: Fluorescence production analysis





- 1) M- DNA Ladder(100-10,000bp),
- 2) S- Bacterial amplified PCR product (850bp)

Figure-5:PCR amplified 16s r RNA

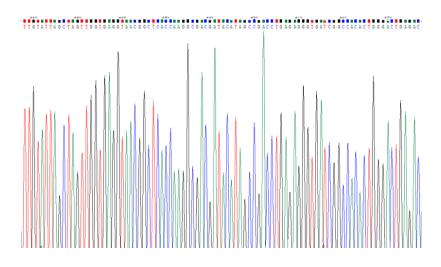


Figure-6: Chromatogram of the sequence

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Job Title	Nucleotide Sequence	Filter Results						
RID	68FZ3SFB013 Search expires on 04-01 17:24 pm Download All ▼	exc						
Program	BLASTN 🚱 Citation 🗸	Organism only top 20 will appear						
Database	nt <u>See details</u> ✓	Type common name, binomial, taxid or group name						
Query ID	Icl Query_44795	+ Add organism						
Description	None	Percent Identity E value	Query Coverage					
Molecule type	dna	to	to					
Query Length	1459							
Other reports	Distance tree of results MSA viewer @		Filter Reset					
Descriptions	Graphic Summary Alignments Taxonomy							
Sequences producing significant alignments Download V New Select columns V Show 100 V								
select all	100 sequences selected	GenBank Graphic	s <u>Distance tree of results</u> New MSA Viewer					
	Description	Scientific Name	Common Name Taxid Max Total Query Score Score Cover					
Pseudomon	as sp. strain ME2020 16S ribosomal RNA gene, partial sequence	Pseudomonas sp.	<u>NA</u> <u>306</u> 1668 2388 88%					
Pseudomon	as fluorescens strain PPA42 16S ribosomal RNA cene partial sequence	Pseudomonas fluorescens	NA 294 1668 2351 87%					

Figure-7:BLAST results

Descriptions	Graphic Summary	Alignments	Taxonomy							
A hover to see the title			Alignment Scor	es 📕 < 40	40 - 50	50 - 80	80 - 200	>= 200	0	
100 sequences selected 💡		Distribution of the top 765 Blast Hits on 100 subject sequences								
			1	250 500	Query 750	1000	1250			

Figure-8:BLAST graphic summary result



Figure-9:Pot assay